The Probiotic Viability Puzzle

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Abstract

Viability is a prerequisite for any therapeutic benefits associated with the ingestion of probiotic bacteria. According to FAO and WHO guidelines, the health-promoting properties of probiotics require that there must be at least 106 colony-forming units per gram of viable probiotic bacteria in the yoghurt at the time of consumption. This project focuses on the challenges in viability determination when probiotic Lacticaseibacillus rhamnosus and Bifidobacterium species are mixed in the yoghurt together with the starter cultures and the technological challenges impacting Bifidobacterium species viability. A quantitative PCR (qPCR) method coupled with an improved version of propidium monoazide (PMAxxTM), and novel species-specific primers was successfully developed for the selective enumeration of the probiotics and starter cultures. The selective quantification of viable cells by PMAxx TM treatment at 100 µM, effectively inhibited and removed >99% of DNA from dead control cells. PMAxxTM-qPCR assay counts were comparable (p > 0.05) to the plate count for all four target organisms. Furthermore, oxidative stress pre-adaptation and high-pressure homogenization were evaluated as potential strategies to overcome the technological challenges impacting probiotic viability during yoghurt processing and storage. Oxidative stress-adapted variants of Bifidobacterium spp. were isolated after exposure to sublethal and lethal H2O2 treatments. The adapted variants were tested for their ability to withstand oxidative stress upon re-exposure (0.1 mM H2O2) using flow cytometry with SYTO 9 and propidium iodide (PI) fluorescent staining to measure membrane integrity as well as Cell ROX green staining to assess the oxidative state. The results suggest pre-adaptation in Bifidobacterium spp. can improve its resilience to oxidative stress, with a better adaptation in B. animalis possibly related to its genetic predisposition. The impact of high-pressure homogenization on the viability of probiotics in full-fat yoghurt under two-stage homogenization at 200, 250, and 600 bar is part of the ongoing work. These innovative approaches for viability optimization and determination would enable inline viability verification and improved probiotic quality assurance for processors.

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Tlaleo Azael Marole is a Ph.D. food science student in the Department of Consumer and Food Sciences at the University of Pretoria, South Africa, under the supervision of Prof. Elna Buys and co-supervisor Dr. Thulani Sibanda. Mr Marole obtained his B.Sc. in Biotechnology at the National University of Lesotho and his M.Sc. in Food Science, Safety and Health with distinction at Heriot-Watt University, Scotland, U.K. Mr. Marole has over six years of work experience in the dairy industry and has worked for various dairy companies in South Africa where he was responsible for the management of quality and food safety systems. Mr. Marole's Ph.D. research focuses on enhancing the survival of probiotic bacteria and the production of bioactive metabolites in a multi-species synbiotic yoghurt during storage.

Ursula Thomashoff graduated with a Bachelor's degree in Food Science and a Bachelor of Science Honours degree from the University of Pretoria. She is currently completing her Master's degree at the same institution. Her expertise lies in Dairy Microbiology and Science. Ursula's groundbreaking research focuses on improving the survival of the probiotic Bifidobacterium spp. during yoghurt manufacturing through stress adaptation techniques. She has already presented her findings at prestigious conferences, including the 2022 ICFMH Food Micro Conference and the 2022 IDF International Conference.

Anthony Hobden is pursuing a master's degree in food science in the department of Consumer and Food Sciences at the University of Pretoria. He holds a BSc in Molecular Biology and Biotechnology from Stellenbosch University and a BSc (Hons) in Microbiology from the University of Pretoria. He has an interest in dairy technology and dairy microbiology.